

Research Article

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Evaluation of Antibacterial Property of Cell-free Hemolymph and Hemocytes of Marine Gastropod, *Rapana Rapiformis* from Inshore Waters of Pondicherry, Southeast Coast of India

S. Amruthalakshmi¹, A. Yogamoorthi^{2*}¹PG Student, Department of Ecology & Environmental Sciences, Pondicherry University, Pondicherry-605014, INDIA²Associate Professor, Department of Ecology & Environmental Sciences, Pondicherry University, Pondicherry-605 014, INDIA

Email: ultrayoga1@gmail.com

ABSTRACT

The cell-free hemolymph and hemocytes isolated from body-fluid of marine gastropod *Rapana rapiformis* distributed in the shallow waters of Pondicherry coast, were tested for their antibiotic potential against five common human bacterial pathogens viz. *Klebsiella pneumonia*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Vibrio cholera* and *E.Coli*. The sensitivity test through Disc diffusion assay revealed that cell-free hemolymph of *Rapana rapiformis* exhibited higher level of inhibition even at MIC of 2.5ul against *Vibrio cholera* when compared to reference drug. A peptide molecule in haemolymph that showed higher inhibitory activity with the molecular mass of 35kDa, has been found by Sodium Dodecyl Sulphate Polyacrylamide gel electrophoresis. It could be of greater interest to isolate and characterize this protein which might be used commercially against existing antibiotic resistant strains such as MRSA in future.

Keywords: Marine gastropod-*Rapana rapiformis*-antibacterial-*vibrio cholera*-low molecular peptide

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INTRODUCTION

Molluscs are widely distributed throughout the world and have many representatives such as slugs, whelks, clams, mussels, gastropods, oysters, scallops, squids and octopods in the marine and estuarine ecosystem (Sharmin Vini et al 2013). Many classes of bioactive compounds exhibiting anti-tumor, anti-leukemic, antibacterial and antiviral activities have been reported worldwide in these group of animals (Rajaganapathy et al., 2000). Among mollusks, some have pronounced pharmacological activities or other properties which are useful in the biomedical area. In addition to the tissue borne secondary metabolites, other components viz. hemolymph and hemocytes are also reported to possess potential bioactive principle across the invertebrates including insects. Molluscan haemocytes stering in the hemolymph are involved with variety of important physiological processes including recognition and elimination of foreign materials, shell repair, wound repair, nutrient distribution and excretion (Armstrong et al 1971; Cheng 1984, Pollero, Huca and Brenuer 1985; Chen and Bayne 1995; Mount et al 2004). Haemocytes which behave like phagocytes are also serve a role in the molluscan stress response involving endocrine molecules (Ottavini, Franchini and Fontalli 1992; Ottavini and Franceshi 1996). Screening studies on antibiotic property and active peptide present in hemolymph and such potential in hemocytes of gastropod body fluid is very meager when compared to studies in insects' hemolymph. Important reports relating to hemolymph bioactive potential in insects viz. Diplopods and Chilopods (Xylander & Neverman,1990, Nevermann et al 1996); crabs viz. *Scylla serrata* (Anitha,

et al 2011); horse shoe crab(Saito et al 1995), *Carcinus menas* (Julit et al 1999;), *Ocypoda macrocera* (Pitchaia Sivaperumal et al 2013) and gastropods viz. *Nerita albicilla*, (Sharmin Vini et al 2013); *Dromia abrohlensis* (Rameshkumar et al 2009) attracted pharmacologist to focus their search for potential secondary metabolites /property in the hemolymph of marine gastropods.

In the present study, an attempt has been made to evaluate the antibacterial property of cell-free haemolymph and hemocytes of the marine gastropod *Rapana rapiformis* against different human pathogens and to ascertain the molecular weight of the protein peptide using SDS-PAGE present in haemolymph responsible for antibacterial property.

MATERIALS AND METHODS

Gastropod, *Rapana rapiformis* is a species of sea snail or marine gastropod mollusk. It is also known as rock snail or murex snail. The colour of the shell is uniform orange-brown; light brown with 3-4 chocolate brown bands on body, the aperture is white with operculum oval in shape. The length of the animal varies from 22.90 to 102.60 mm, diameter from 17.90 to 84.76 mm. The max height of the aperture is 9.60 to 83.75 mm and height of spire is 8.49 to 33.68 mm. Live animals are collected alive from fishing launches operating in the inshore waters of Pondicherry, southeast coast of south India.

Collection of haemolymph & isolation of hemocytes

The haemolymph was obtained from the opercular region between the shell and mantle tissue using sterile 5ml syringe and needle. Totally 12ml of haemolymph was collected from 6 animals. The haemolymph was centrifuged at 10000rpm for 30 minutes to separate hemocytes. The supernatant was collected and stored at -4°C. The hemocytes were also separated and stored in the same way for further use (Fig.1a &1b).



Figure 1a: *Rapana rapiformis*



Figure 1b: Collection of Haemolymph

Antibacterial assay

The bacterial strains used for sensitivity test were *Klebsiella pneumonia*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Vibrio cholera* and *E.Coli* obtained from the stock available in the department.

Disc Diffusion Method

The antibacterial activity of cell-free haemolymph and hemocytes collected marine gastropod *Rapana rapiformis* was against five strains viz, *Klebsiella pneumonia*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Vibrio cholera* and *E.Coli*. Nutrient broth medium (MullerHinton agar) was prepared and sterilized at 121lbs for 15minutes. In vitro antibacterial assay were conducted using the standard disk-diffusion method (McCaffrey and Endean., 1985). 20uL of the sample was impregnated to what man No.1 filter paper. Disks with standard antibiotics (Erythromycin and Ampicillin) were also prepared and allowed to dry at room temperature. Further they were aseptically placed on the agar plates swabbed with the test microorganisms. The plates were incubated at 37°C for 24hours. The antibacterial activity was measured accordingly based on the zone of inhibition around the disk impregnated with haemolymph. (Sharmin Vini et al 2013) using standard scale obtained from Hi-media.

SODIUM DODECYL SULFATE-POLY ACRYLAMIDE GEL ELECTROPHORESIS

One-dimensional SDS-PAGE was performed using standard methods on the Bio-Rad Mini-Protean II system. It is discontinuous system with 12% separating gel(pH 6.8) of size 7cm x 10cm x 1cm. Prior to electrophoresis, protein samples were redissolved in Laemmli buffer (Laemmli, 1970) and boiled in the presence of dithiothreitol (DDT) for 5min at 100°C. For the molecular weight markers, Sigma marker wide molecular weight range (sigma) was used. The electrophoresis was performed at 10Ma per gel. The gels were stained with Brilliant Blue R concentrate (Sigma-

Aldrich) for 30mins and were destained in 50 % (v/v) methanol, 5 % (v/v) acetic acid for 30mins or until band appeared.

RESULTS

Antibacterial activity study

The antibacterial activity of haemolymph and haemocytes of *Rapana rapiformis* was measured as radius of zone of inhibition (diameter in mm) around the disc in comparison with the standard drugs Ampicilin and Erythromycin. The haemolymph of *Rapana rapiformis* showed significant inhibitory effect against human pathogenic bacteria *Vibrio cholerae*, whereas haemocytes showed higher activity against *Vibrio cholerae*, *Psuedomonas aeruginosa* and *Escherichia coli* (Fig.2). The haemolymph was ineffective against the other four human bacterial pathogens such as *Psuedomonas aeruginosa*, *Klebsiella pneumonia*, *Escherichia coli*, and *Staphylococcus aureus* and haemocytes were ineffective against *Staphylococcus aureus* and *Klebsiella pneumonia* as shown in Tables 1&2 and Figs. 2&3 respectively. The minimum inhibition concentration (MIC) for haemolymph against the pathogen *Vibrio cholerae* was obtained as 2.5ul whereas concentration of haemocytes against *Vibrio cholerae* is 5.0ul . *Escherichia coli* and their MIC values for both was 5.0ul.

Table: 1 Antibacterial activity of Haemolymph of *Rapana rapiformis* against different human pathogen

| Tested pathogens | Zone of inhibition(mm) | | |
|-------------------------------|------------------------|------------------------|---------------------|
| | Haemolymph (mm) | Reference Drugs (mm) | |
| | 20µl /disc | Erythromycin (10ug/ml) | Ampicilin (10ug/ml) |
| <i>Vibrio cholera</i> | 26 | 27 | 25 |
| <i>Klebsiella pneumonia</i> | Nil | 26 | 22 |
| <i>Staphylococcus aureus</i> | Nil | 27 | 25 |
| <i>Psuedomonas aeruginosa</i> | Nil | 26 | 24 |
| <i>E. coli</i> | Nil | 24 | 26 |

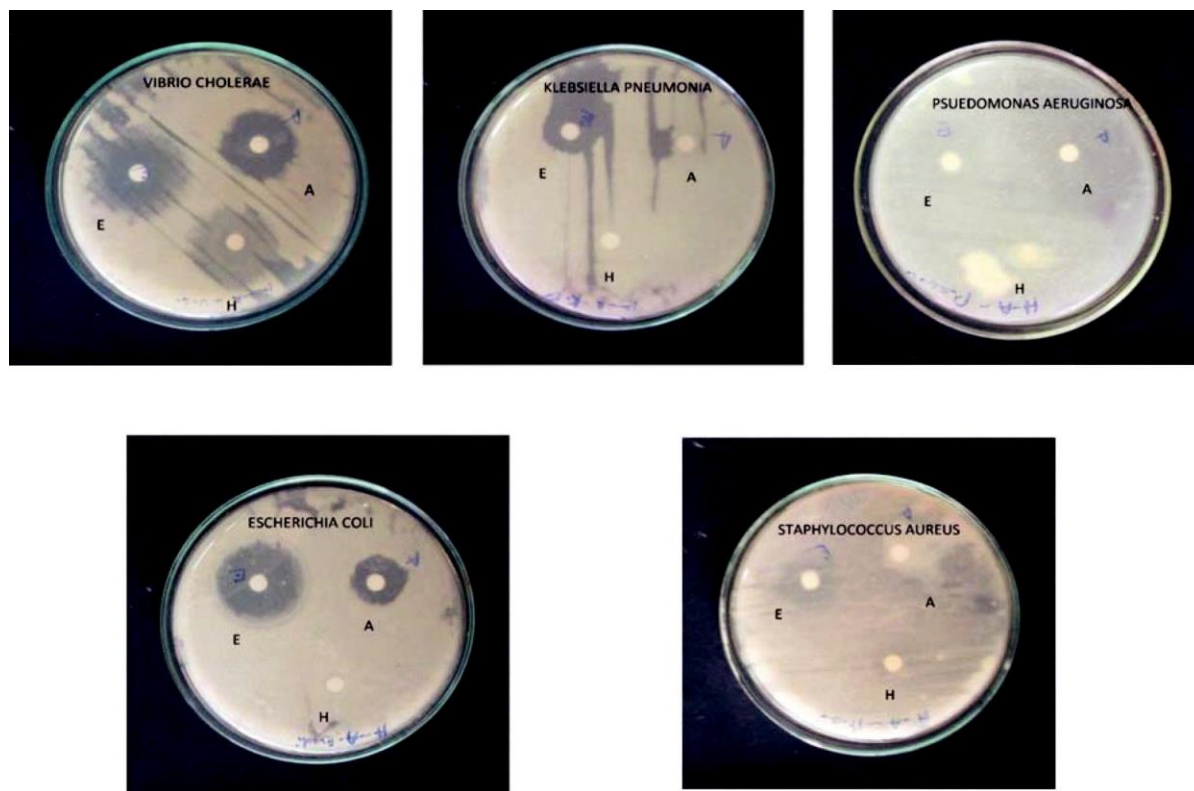


Figure 2: Antibacterial activity of Haemolymph of *Rapana rapiformis* against different Human pathogens E – Erythromycin A- Ampicilin H- Haemolymph

Table 2: Antibacterial activities of Haemocytes of *Rapana rapiformis* against different human pathogens

| Tested pathogens | Zone of inhibition(mm) | | |
|-------------------------------|------------------------|---------------------|---------------------|
| | Haemoytes (mm) | Reference Drugs(mm) | |
| | 20 μ l /disc | | Ampicilin (10ug/ml) |
| <i>Vibrio cholera</i> | 20 | 28 | 18 |
| <i>Klebsiella pneumonia</i> | Nil | 14 | 18 |
| <i>Staphylococcus aureus</i> | Nil | 15 | 21 |
| <i>Psuedomonas aeruginosa</i> | 25 | 32 | 25 |
| <i>E. coli</i> | 19 | 31 | 21 |

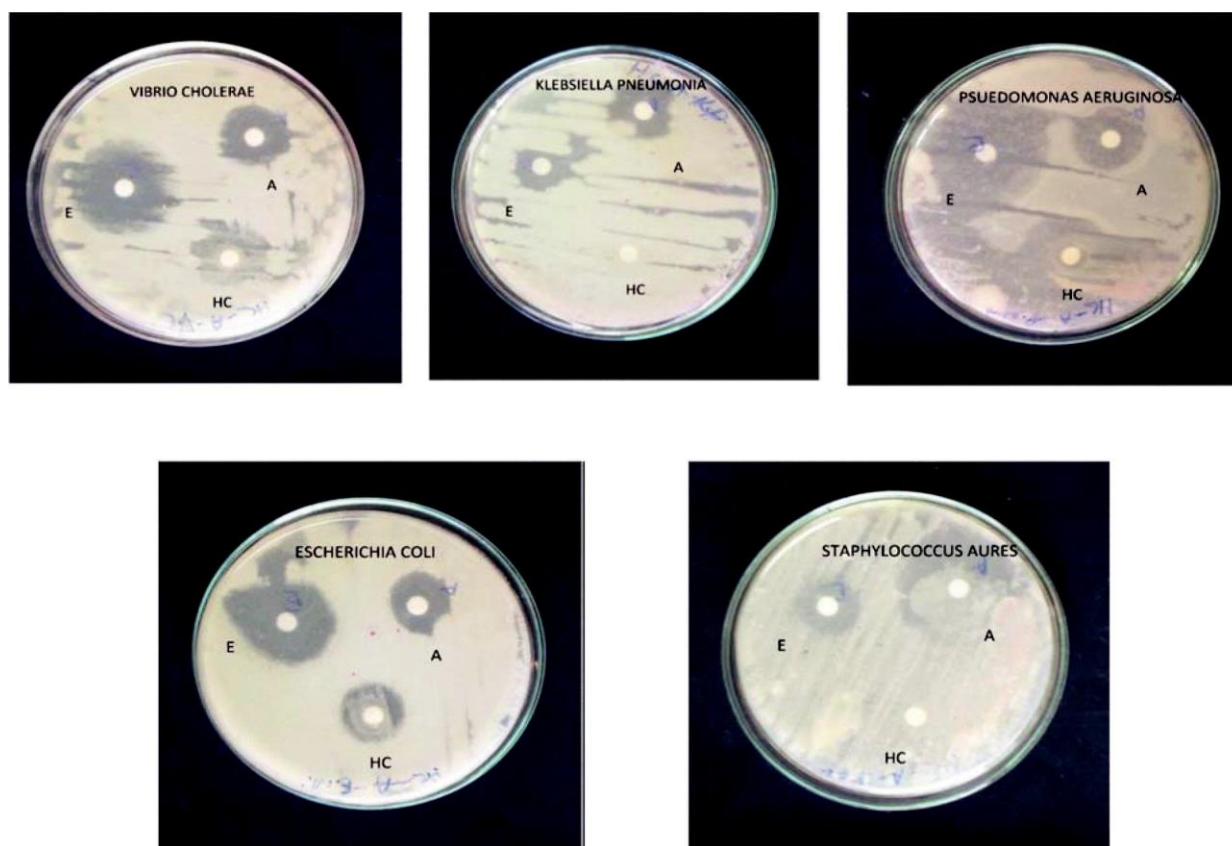
Figure 3: Antibacterial activity of Haemocytes of *Rapana rapiformis* against different human pathogens E- Erythromycin A- Ampicilin H- Haemocytes.

Table 3: Minimum Inhibition Concentrations (MIC)

| Name of the pathogen | Concentration of Haemolymph (ul) | Concentration of Haemocytes (ul) |
|-------------------------------|----------------------------------|----------------------------------|
| <i>Vibrio cholera</i> | 2.5 | 5.0 |
| <i>Klebsiella pneumonia</i> | No activity | No activity |
| <i>Psuedomonas aeruginosa</i> | No activity | 5.0 |
| <i>Staphylococcus aureus</i> | No activity | No activity |
| <i>Escherichia coli</i> | No activity | 5.0 |

SDS GEL Electrophoresis

A peptide molecule in haemolymph with the molecular mass of 35kDa has been found by Sodium Dodecyl Sulphate Polyacrylamide gel electrophoresis

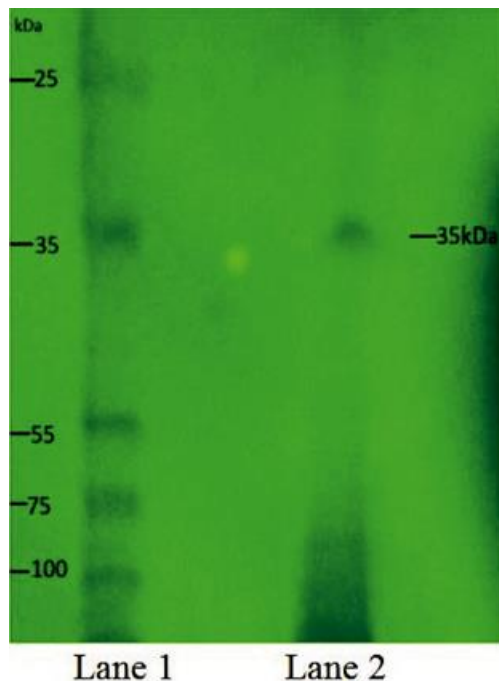


Figure 4: SDS PAGE for hemolymph

Lane 1- Standard protein marker Lane 2- Sample (Haemolymph)

DISCUSSION

The emergence of new infectious diseases and development of resistance to the antibiotics by the existing ones led to search for new but better potent source of antibiotic. This field of research receives the attention of investigators from various fields such as marine biology, marine microbial ecology, biochemistry, chemistry, pharmacology and biotechnology. In the industrialized countries, about 25% of all prescribed drugs contain active principles that are still extracted from higher plants. Consequently there is growing interest in marine natural products or marine-borne secondary metabolites (Sharmin Vini et al. 2013). Human studies on antibacterial property of haemolymph of gastropods are very meager. In the present study hemolymph and haemocytes isolated from marine gastropod *Rapana rapiformis*, have been evaluated for its antibacterial property. *Vibrio cholerae* have shown higher sensitivity among the tested pathogens with a zone of inhibition of 26mm. And the test of hemocytes revealed that hemocytes have shown activity against three human pathogens such as *Vibrio cholerae*, *Pseudomonas aeruginosa* and *Escherichia coli* with a zone of inhibition of 20mm, 25mm, and 21mm respectively. Similar observation was made by Sharmin Vini et al. (2013) where the antimicrobial activity the zone of inhibition 8mm was recorded against *B. subtilis*, *K. pneumoniae* and *S. typhi*, by the hemolymph of gastropod *Nerita albicilla* and the similar trend was also recorded against *E. coli*, *K. pneumoniae* and *S. aureus*, by the hemolymph of *Purpura bufo*, another marine gastropod (Sharmin Vini et al. (2013). Further an attempt was made to ascertain the active peptide present in the haemolymph of *Rapana rapiformis* through the SDS-PAGE revealed that the hemolymph *Rapana rapiformis* possess a protein with molecular mass of 35kDa. Similar protein was also found in hemolymph of shore crab *Ocypoda macrocera* MW ranging between 15.43 to 60.34kDa. (Pitchiah Sivaperumal et al., 2013). A 27kDa protein was purified from horse-shoe crab hemocyte (Saito et al., 1995.). Another antibacterial protein of 11.5kDa was purified and characterised from *Carcinus maenas* which is cationic and hydrophobic in nature and showed inhibition only against marine gram-positive bacteria (Juliet et al., 1999). Protein estimation in cell free and precipitated haemolymph of crab species *Scylla serrata* and *Metagrapsus messor* showed remarkable variations. SDS-PAGE showed many protein bands with molecular size ranging from 22 to 60KDa. Similarly SDS-PAGE of haemocyte contents showed protein bands with molecular size ranging from 22 to 91KDa was reported. (Samuthirapandian et al., (2010). Likewise Pan et al., (2008) reported two bands at molecular weight around 73 and 75 KDa from haemocyte *Bombyx mori*. Low molecular peptides were also isolated from the haemolymph of various insects. The haemolymph of diplopod as a reference showed that its lysozyme had a molecular weight of 14.5kDa (Xylander et al 1997). From chilopod haemolymph low molecular peptides ranging from 15.5 to 16.5kDa was isolated by Xylander (2009). Reports on haemolymph antibiotic peptides also emphasize that in insects which lack an adaptive immune system, antimicrobial peptides play a crucial role in fighting invading pathogens. They are synthesized in response to microbial infection or septic body injury mainly in insect

fat body (functional equivalent of mammalian liver) and in certain blood cells, and then rapidly released into hemolymph where they act synergistically against microorganisms. (Tzou et al., 2002; Dunphy et al., 2003 and Irving et al., 2004). Similarly, it is presumed that circulating haemolymph with hemocytes in marine invertebrates contains biologically active substances such as complement, lectins, clotting factors, antimicrobial peptides and lipids such as fatty alcohols, free fatty acids and monoglycerides (Miyata et al,1989;Thomar et al.,2007). Therefore, it could be of greater interest to isolate and characterize this protein which might be used commercially against those resistant strains because it has become a normal feature that bacteria are developing resistance to antibiotic after several uses. Concludingly, based on present findings on the antibiotic peptides present in the hemolymph and reports on various invertebrates particularly in gastropods, it could be stated that body fluid of invertebrates i.e. hemolymph and streaming hemocytes are found to be potential source of antibiotic and further confirmation, isolation purification and characterization of those active peptides would help to combat antibacterial resistant microbial pathogenicity like Methicillin resistant Staphylococcus aureus (MRSA).

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